# Stress Alterations in Growth Parameters, Pigment Content and Photosynthetic Functions of *in vitro* Cultured Plants

Judit Kissimon<sup>a,\*</sup>, Ágnes Tantos<sup>a</sup>, Annamária Mészáros<sup>a</sup>, Erzsébet Jámbor-Benczúr<sup>b</sup>, Gábor Horváth<sup>a</sup>

a Department of Plant Physiology and

- Department of Floriculture and Dendrology, University of Horticulture and Food Industry, P. O.Box 53, Budapest, Hungary, H-1518. Fax: 00-36-1209-6388.
  E-mail: KISSIMON@HOYA.KEE.HU
- \* Author for correspondence and reprint requests
- Z. Naturforsch. 54c, 834-839 (1999); received November 28, 1998/March 18, 1999

Anthocyanin, Fluorescence, Huckberry, Peach, Triacontanol

Effects of different concentrations of glucose, sucrose and the natural growth regulator, triacontanol were studied under the unfavourable stress conditions of micropropagation of two woody plants, Sorbus rotundifolia L. and Prunus × davidiopersica 'Piroska'. After 4–6 weeks of cultures, the number and length of shoots, the photosynthetic activity as well as the chlorophyll, carotenoid and anthocyanin contents were investigated. As shown by the growth parameters, the optimal carbohydrate concentration was between 1.5–2.5%, whereas in higher concentrations, a definite inhibition could be observed. A similar response was found in changes of the anthocyanin content in Prunus × davidiopersica 'Piroska', but this effect was less pronounced with the photosynthetic pigments in both species. The Fv/Fm ratio representing the quantum yield of photosynthesis was low due to the inhibitory effect of stress and altered significantly by changing the carbohydrate concentrations. In all cases, the addition of  $2-4\,\mu g$  triacontanol/l further enhanced the stimulating effect of the optimal carbohydrate concentrations, which indicated the specific importance of the appropriate hormone balance under such stress conditions.

# Introduction

The artificial environment of micropropagation (including the mechanical injury, changes of the nutrition and hormone availability, light intensity and quality, temperature, etc.) causes dramatic stress for developing plantlets. Amongst various factors, the concentration of carbohydrates and growth regulators in the culture medium play an important role in affecting the development of plantlets. For minimizing the stress effects induced by the artificial environment, optimization of the carbon source and hormone availability is specifically required.

Even sucrose or glucose, the most frequently used carbon sources have a retarding effect on growth in unfavorable concentrations. In tissue culture media for woody plants sucrose is usually used in a concentration of 3% (George, 1993), a rate that was originally recommended by Murashige and Skoog (1962) for callus culture of *Nicotiana*. In other studies the effect of various monosaccharides on the proliferation of woody plants

was investigated and the "magic" 3% or very similar concentrations of carbohydrate source were applied as well (Belaizi and Boxus, 1995; Druart, 1995; Karhu, 1995). The effect of carbohydrates above or below the generally used concentration was only studied by Karhu and Ulvinen (1995) and Jámbor-Benczúr *et al.* (1996).

In micropropagation of woody plants, artificial growth regulators (benzyladenine, 2-isopentenyladenine) are generally used in the multiplication phase (Einset, 1991). Zeatin, a natural cytokinin would be the most favorable for plants, but it is seldom used as it is not economical. Therefore, it is of great importance to find natural growth regulators, which can be used to make mass propagation economical. Triacontanol, a long-chain primary alcohol which is a natural component of waxes was found to stimulate plant growth in vivo (Steffens and Worley, 1980; Ries, 1985; Srivastava and Sharma, 1991). Very recent work demonstrated that addition of triacontanol to the in vitro culture medium of Melissa officinalis resulted in high proliferation and significantly increased the

0939 – 5075/99/0900 – 0834 \$ 06.00  $\,$  © 1999 Verlag der Zeitschrift für Naturforschung, Tübingen  $\cdot$  www.znaturforsch.com  $\cdot$  D



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

Zum 01.01.2015 ist eine Anpassung der Lizenzbedingungen (Entfall der Creative Commons Lizenzbedingung "Keine Bearbeitung") beabsichtigt, um eine Nachnutzung auch im Rahmen zukünftiger wissenschaftlicher Nutzungsformen zu ermöglichen.

On 01.01.2015 it is planned to change the License Conditions (the removal of the Creative Commons License condition "no derivative works"). This is to allow reuse in the area of future scientific usage.

chlorophyll content of the plantlets (Tantos *et al.*, 1999). Since the induction of proliferation in woody plants is difficult in most cases and the plantlets are usually pale due to the low pigment content induced by the stress condition of *in vitro* culture, triacontanol seemed to be a useful tool to overcome the above mentioned problems.

The aim of this work was therefore to optimize the concentrations of both carbon sources and triacontanol as a natural growth regulator in the micropropagation of two woody plants by monitoring alterations in their growth parameters, in their pigment content and in the photosynthetic activity characterized by fluorescence induction kinetics. Our data have clearly demonstrated that the usually applied micropropagation conditions are far from the optimum required for minimizing the harmful effect of the artificial environment.

#### **Materials and Methods**

Plant materials and growth conditions

In vitro cultures of a roadside tree, huckberry (Sorbus rotundifolia L. cv. 'Bükk Szépe'), and an ornamental peach tree with red leaves (Prunus × davidiopersica 'Piroska') were investigated for two years. The basal medium for S. rotundifolia 'Bükk Szépe' was MS medium (Murashige and Skoog, 1962) supplemented with hormones according to Molnár et al. (1994). Prunus × davidiopersica 'Piroska' was cultured on S-basal medium supplemented with 0.75 mg BA-ribose/l, 0.1 mg IBA/l and 0.1 mg GA<sub>3</sub>/l (Jámbor-Benczúr and Márta-Riffer, 1990). Sucrose or glucose was added at 0.5 to 3% concentrations into the culture media of S. rotundifolia or Prunus × davidiopersica 'Piroska'. The combined effect of carbohydrates and triacontanol was studied on Prunus × davidiopersica 'Piroska' by supplementing the sugar-containing media with 2, 4 or 8 µg triacontanol/l. The in vitro cultures of both plants were illuminated with white light (Philips TLD 30W/827, F30W/33 and Tungsram 30W normal) at the intensity of 40 µmol·m<sup>-2</sup> ·s<sup>-1</sup> by using 16/8 hours light/dark cycles with the day/night temperature of 22/16 °C, respectively.

## Growth analysis

The number and length of the newly initiated shootlets of both S. rotundifolia and  $Prunus \times da$ -

vidiopersica 'Piroska' were counted on 8 explants of each treatment in 4 replications.

## Determination of pigment contents

Chlorophyll (a+b) content of each plant was estimated in 80% acetone extracts as described by Arnon (1949). The concentration of carotenoids was determined according to Horváth *et al.* (1972). The anthocyanin content was measured in shoots of *Prunus* × *davidiopersica* 'Piroska' according to the method of Kho *et al.* (1975).

#### Photosynthesis measurements

Chlorophyll fluorescence induction kinetics were measured on huckberry plants by using a portable fluorimeter (Plant Efficiency Analyser, Hansatech). After 20 minutes of dark adaptation samples were excited with the actinic light 1800 µmol·m<sup>-2</sup>·s<sup>-1</sup> of intensity at 650 nm. Fluorescence was detected for 60 s and the usually used parameter, the Fv/Fm ratio was calculated (Bolhár-Nordenkampf and Öquist, 1993). Six leaves freshly collected from a *Sorbus rotundifolia* L. cv. 'Bükk Szépe' tree in the Garden of the University were used as control samples.

Student's t-test was used for statistical evaluation of experimental data. The number of data (n), standard deviation and the level of significance are presented in the figures.

#### **Results and Discussion**

The various concentrations of different carbohydrates added to the culture medium strongly influenced the proliferation ability of S. rotundifolia shoots (Fig. 1). For either sucrose or glucose, the lowest and highest sugar concentrations yielded fewer shoots. Each concentration of glucose resulted in better sprouting compared to similar levels of sucrose. The highest shoot numbers were obtained at 1.5% glucose and 2% sucrose concentrations. As compared to the lowest sugar concentration longer shoots developed on the media containing 1.5 to 2.5% of both carbohydrates and even at 3% of glucose. Similar results were obtained with Prunus × davidopersica 'Piroska' where the optimal concentration of both carbohydrates was 2% (Fig. 2). In earlier work dealing with the micropropagation of different Rosaceae

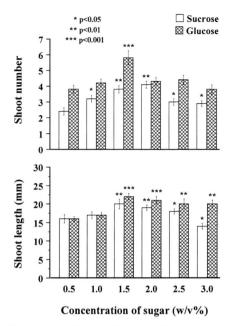


Fig. 1. Influence of increasing carbohydrate concentration of the culture media on the shoot number and length of micropropagated *Sorbus rotundifolia* (n = 32, stars indicate the level of significance at different sugar concentrations at 0.05, 0.01 and 0.001 probability).

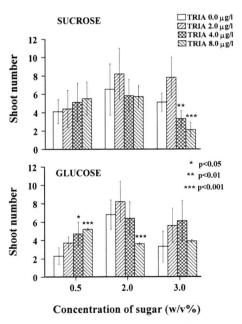


Fig. 2. Comparison of the triacontanol effect at different concentrations on the shoot number of  $Prunus \times davidi-opersica$  'Piroska' (n = 32, stars indicate the level of significance at different triacontanol concentrations at 0.05, 0.01 and 0.001 probability).

comparing the effect of various carbohydrates, the sugar concentration was generally 3% (Druart, 1995; Karhu, 1995). Our results have clearly demonstrated that the sugar concentration used earlier for the micropropagation of *Sorbus sp.* is far from the optimum and perhaps can rather be considered as a stress agent. This conclusion agrees with the results of Molnár *et al.* (1994) and Jámbor-Benczúr *et al.* (1995) where 2% sugar was found to be the most suitable concentration for micropropagation of *S. rotundifolia* and *Prunus* × *davidopersica* 'Piroska', and higher concentrations of both sugars harmful by affecting the plantlets.

The effect of triacontanol combined with various carbohydrate concentrations is also shown in Fig. 2. At 0.5% of carbohydrates, all triacontanol concentrations applied resulted in higher yields in shoot proliferation. The highest shoot number was obtained with 2 µg/l triacontanol at 2% sugar and further elevation of the sugar or triacontanol concentrations inhibited proliferation. This agrees with an earlier finding where 2 µg triacontanol/l was found to be optimal for the micropropagation of *Melissa officinalis* (Tantos *et al.*, 1999).

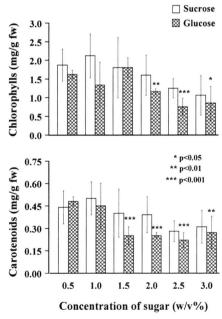


Fig. 3. Influence of different sucrose and glucose concentrations of the culture media on the photosynthetic pigment content of micropropagated *Sorbus rotundifolia* shootlets (n = 8, stars indicate the level of significance at different sugar concentrations at 0.05, 0.01 and 0.001 probability).

Photosynthetic pigment analysis of huckberry plantlets grown at different sugar concentrations indicated that with the exception of the very low concentrations (0.5-1.0%), higher amounts of sucrose or glucose in the culture media led to significant less the chlorophyll or carotenoid content of the leaves (Fig. 3). At most sugar concentrations, the chlorophyll content of plantlets was higher on sucrose than on glucose, but the high sucrose concentrations were also inhibitory to proliferation. This result is in agreement with Donelly and Vidaver (1984) who found less pigment content and photosynthesis at higher concentration of sugar in red raspberry in vitro and ex vitro cultures. Our results prove that high concentrations of both sugars can be considered as stress factors, which is very probably related to their osmotic effect (Sato et al., 1996).

The combined effect of sugars and triacontanol on the chlorophyll content of shootlets of *Prunus* × *davidopersica* 'Piroska' is shown in Fig. 4. The effect of sucrose and glucose was basically similar to the result obtained with *S. rotundifolia* (c.f. Fig. 3 and Fig. 4). The lower concentrations (2 to

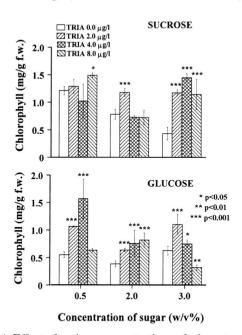


Fig. 4. Effect of various concentrations of triacontanol of the culture media on the accumulation of chlorophylls in micropropagated  $Prunus \times davidiopersica$  'Piroska'  $(n = 8, \text{ stars indicate the level of significance at different triacontanol concentrations at 0.05, 0.01 and 0.001 probability).$ 

4 μg/l) of triacontanol could overcome the inhibitory effect of the higher sucrose concentrations. The high concentrations of triacontanol were also found to be inhibitory similar to results previously described by Tantos *et al.* (1999). The inductive effect was less pronounced at 0.5% sucrose concentrations but at 2% sucrose 2 μg triacontanol/l produced significantly higher chlorophyll content. Applying triacontanol together with glucose, however, enhanced the chlorophyll content at each glucose concentration.

Photosynthetic efficiency is usually characterized by the quantum yield of photosynthesis represented by the ratio of the maximal (Fm) and variable (Fv) components of the fluorescence induction curve. Under normal conditions, the Fv/Fm ratio is around  $0.832\pm0.004$  and its decrease is a good indicator of the manifestation of stress (Bolhár-Nordenkampf and Öquist, 1993). Regarding the Fv/Fm ratio of *in vitro* cultured *S. rotundifolia* shoots, we found significant lower Fv/Fm values as compared to that of the control leaf (Fv/Fm = 0.838). No differences were found at different glucose concentrations but it was strongly varied with changing the sucrose concentration of the medium. The overall decrease of the Fv/Fm

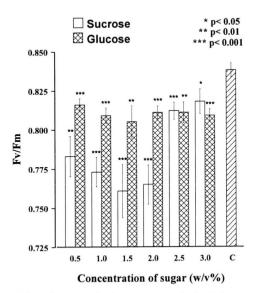


Fig. 5. The photosynthetic quantum yield of *S. rotundifolia* at various sugar concentrations as measured by chlorophyll fluorescence induction. (C – control tree leaves of the Garden, n = 12, stars indicate the level of significance at different sugar concentrations at 0.05, 0.01 and 0.001 probability).

ratio indicated the general stress effect of micropropagation (Fig. 5). Van Huylenbroeck and Debergh (1996) also experienced a 10% decrease in the Fv/Fm ratio and the photosynthetic pigment content at the beginning of sucrose treatment but after 14 days of the acclimation, the photosynthesis recovered and the pigment content increased. In contrast, Karhu and Ulvinen (1995) measured practically no decline in the Fv/Fm ratio in various sucrose media as compared to other carbohydrate treatments.

Prunus × davidiopersica 'Piroska' contains anthocyanin, the red pigment in its leaves. In general the increasing anthocyanin content in leaves is considered as an indicator of the appearance of stress because the activity of phenylalanine ammonia lyase increases under unfavorable conditions (Tholakalabavi et al., 1997). Since anthocyanin is a natural constituent of Prunus × davidiopersica 'Piroska', the presence of red color does not mean a disadvantage for the plantlets. As shown in Fig. 6 the increasing concentrations of carbohydrates enhanced the anthocyanin accumulation in leaves which peaked at 2% of both sugars mainly in glu-

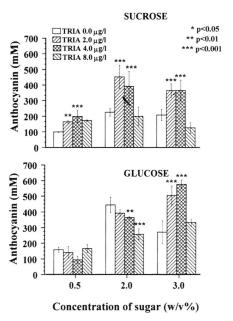


Fig. 6. Combined effect of different concentrations of sugars and triacontanol of the culture media on the anthocyanin accumulation in micropropagated *Prunus*  $\times$  *davidiopersica* 'Piroska' (n = 8, stars indicate the level of significance at different triacontanol concentrations at 0.05, 0.01 and 0.001 probability).

cose. The stimulating effect of triacontanol is more pronounced in the presence of sucrose than glucose but the highest triacontanol concentration had an inhibitory effect similar to that observed with chlorophylls, shoot number and length. We assume, therefore, that the elevated anthocyanin content at the optimal concentration of carbohydrates and triacontanol represents the restoration of the normal anthocyanin synthesis, rather than a manifestation of unfavourable stress conditions.

This work demonstrates that for *Sorbus rotundifolia* and *Prunus* × *davidiopersica* 'Piroska' the inhibitory effect of stress induced by micropropagation can be reduced by optimizing the concentration of carbohydrate source and the conditions can further be improved by adding triacontanol to the culture.

### Acknowledgement

This work was partially supported by the Hungarian National Scientific Research Fund (OTKA T-017686, T-026078).

- Arnon P. I. (1949), Copper enzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris*. Plant Physiol. **24**, 1–15.
- Belaizi M. and Boxus P. (1995), *In vitro* shoot multiplication of cork oak (*Quercus suber* L.). Influence of different carbohydrates. Bull. Rech. Agron. Gembloux **30**, 39–46.
- Bolhár-Nordenkampf H. E. and Öquist G. (1993), Chlorophyll fluorescence as a tool in photosynthesis research. In: Photosynthesis and Production in a Changing Environment (Hall, D. O., Scurlock, J. M. O., Bolhár-Nordenkampf, H. R., Leegood, R. C. and Long, S. P., eds.). Chapman & Hall, London, pp. 193–206.
- Donelly D. J. and Vidaver W. E. (1984), Leaf anatomy of red raspberry transferred from culture to soil. Amer. Soc. Hort. Sci. **109**, 177–181.
- Druart P. (1995), C-source and growth response of *Prunus glandulosa* 'Sinensis' THUND. and *Malus pumila* MILL. M26 and M9 clone 29 during *in vitro* propagation. Bull. Rech. Agron. Gembloux **30**, 29–38.
- Einset J. W. (1991), Woody plant micropropagation with cytokinins. In: High-Technology and Micropropagation I. Biotechnology in Agriculture and Forestry (Bajaj Y. P. S. ed.). Springer Publ. Vol. 17, pp. 190–201.

George E. R. (1993), Plant Propagation by Tissue Culture. Exegetics Lim., Edington, England.

- Horváth G., Kissimon J. and Faludi-Dániel Á. (1972), Effect of light intensity on carotenoids in normal and mutant leaves. Phytochemistry 11, 183–187.
- Jámbor-Benczúr E. and Márta-Riffer, A. (1990), In vitro propagation of *Philodendron tuxtlanum* Bunting with benzylaminopurine. Acta Agronomica Hungarica 39, 341–348.
- Jámbor-Benczúr E., Tilly-Mándy A. and Szafián Zs. (1995), Effect of growth regulators on in vitro propagation of Prunus × davidiopersica 'Piroska'. IX. Forum for Applied Biotechnology, Med. Fac. Landbouw Univ. Belgium 60, 1665–1668.
- Jámbor-Benczúr E., Onadi K., Kissimon J. and Horváth G. (1996), The effect of carbon source on *in vitro* multiplication, photosynthesis and anatomical structure of *Sorbus rotundifolia* L. Acta Horticult. **447**, 157–159.

- Karhu S. T. (1995), The quality of applied carbohydrates affects the axillary branching of apple microshoots. Bull. Rech. Agron. Gembloux **30**, 21–27.
- Karhu S. T. and Ulvinen S. K. (1995), The effect of different carbohydrates on the rooting of micropropagated apple shoots and their adaptation after transplantation. Bull. Rech. Agron. Gembloux **30**, 87–101.
- Kho K., Bennink G. J. and Wiering H. (1975), Anthocyanin synthesis in a white flowering mutant of *Petunia hybrida* by complementation technique. Planta **127**, 271–279.
- Molnár K., Mándy A., Jámbor-Benczúr E. and Márta K. (1994), Preliminary results in micropropagation of *Sorbus rotundifolia* L. Public. Univ. Hort. Ind. Aliment. **54**, 106–108.
- Murashige T. and Skoog F. (1962), A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plant. **15**, 473–479.
- Ries S. K. (1985), Regulation of plant growth with triacontanol. Crit. Rev. Plant Sci. 3, 239–251.
- Sato K., Nakayama M. and Shigeta J. (1996) Culturing conditions affecting the production of anthocyanin in suspended cell cultures of strawberry. Plant Sci. Limerick. **113**, 91–98.
- Srivastava N. K. and Sharma S. (1991), Effect of triacontanol on photosynthesis, alcoholic content and growth in opium poppy (*Papaver somniferum*). Plant Growth Regulators **9**, 65–71.
- Steffens G. L. and Worley J. F. (1980), Triacontanol. Evolution in Several Plant Assays. Proc. 7th Ann. Plant Growth Regulator Working Group, Dallas, July, 13–15.
- Tantos A., Mészáros A., Kissimon J., Horváth G. and Farkas T. (1999), The effect of triacontanol on micropropagation of balm, *Melissa officinalis L.* Plant Cell Report (in press).
- Tholakalabavi A., Źwiazek J. J. and Thorpe T. A. (1997), Osmotically-stressed poplar cell cultures: anthocyanin accumulation, deaminase activity, and solute composition. J. Plant Physiol. **151**, 489–496.
- Van Huylenbroeck J. M. and Debergh P. C. (1996), Impact of sugar concentration *in vitro* on photosynthesis and carbon metabolism during *ex vitro* acclimatisation of *Spathiphyllum* plantlets. Physiol. Plant. **96**, 293–304.